

## **MYCOPROSAFE®**

### **QUICK APPLICATION GUIDE**

**WARNING:** Use only conical centrifuge holders of 30 mm of diameter adapted to the shape of the sampling tubes

#### **Sputum and samples other than urine:**

1. Transfer a maximum volume of 10 mL of sample from the collection cup into the tube containing the glass beads.
2. Add Sodium Hydroxide - Sodium Citrate solution in a quantity approximately equal to that of the sample. Close the cap tightly.
3. Homogenize the sample by shaking with a Vortex and incubate from 15 to 30 minutes at room temperature.
4. Fill the tube to the "50 mL" line with phosphate buffer solution (save the left over buffer for further steps).
5. Spin the tubes for 15 minutes at 2000 x g.
6. Carefully discard the supernatant into a container with disinfectant solution. Leave as little solution as possible in the tube.
7. Add a few milliliters of the remaining phosphate buffer but not above the 5 mL line. Do not fill past the 5 mL line. Resuspend the sediment left between the glass beads, by vortexing. This suspension can now be used for culture, microscopy and molecular diagnostic methods.
8. Once the procedure is complete, disinfect the tubes using the disinfectant solution provided in the kit before discarding.

#### **Urine samples:**

It is recommended to obtain early morning urine to increase the chance of recovery of mycobacteria.

1. Transfer the urine samples from the collection cup into the tube that contains the glass beads. The tube can be filled up to the "50 mL" line.
2. Spin the tubes for 15 minutes at 2000 x g.
3. Discard the supernatant into a container with disinfectant solution. Add approximately 3 mL of sodium hydroxide-sodium citrate solution onto the glass beads. Close the cap securely.
4. Continue processing, by starting from step 3 described above for other types of samples.